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The Toxicity of *n*-Hexane, Ethyl Acetate and Ethanol Extracts of SarangBanua (Clerodendrumfragrans Vent Willd) Leaves by Brine Shrimp Lethality Test (BSLT) Method

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Abstract.Sarangbanua traditional medicinal plant is found in Simalungunand NorthTapanuli Regencies, Sumatera, Indonesia. The result of plant determination, sarangbanuais Clerodendrumfragrans Vent Willd, including the Verbenaceae family. This study aims to determine the toxicity of the three types of leaf extracts of C. fragrans Vent Will. The Brine Shrimp Lethality Test (BSLT) method was used to determine the toxicity of the extracts. The mortality data were then analyzed by Probit SAS to obtain LC50 values. The results showed that the three types of C. fragrans leaf extracts had LC50 values below 1000 µg/mL, so they were classified as toxic and potentially bioactive. The LC50 values of each of the ethanol, ethyl acetate and ethanol extracts were 26.25; 37.50 and 41.97 µg/mLKeywords: Clerodendrumfragrans Vent Willd, toxicity, brine shrimp lethality test (BSLT), Indonesian medicinal plants.

1. Introduction

The high biodiversity of Indonesian plant has the potential to be developed as raw material for natural medicine. One of the plants that have been used by the people of the Simalungun area, Sumatera as traditional medicine is the sarangbanua. Sarangbanuais Clerodendrumfragrans Vent Willd, including the Verbenaceae family according to Herbarium Botani LIPI Cibinong, in June, 2017. This plant has been used as a traditional medicinal plant for stomach aches, fever, high blood pressure medicine, and diabetes.

Secondary metabolites alkaloids, triterpenoids, flavonoids, saponins, tannins, quinones are present in the ethanol extract of C. fragrans leaves. The alkaloids, steroids, saponins, tannins are present in ethylacetate extracts and in the extract *n*-hexane there are alkaloids, triterpenoids, flavonoids [1]. These secondary metabolites found in C. fragransare potential for these plants to become medicinal plants. Another plant same family is С in the the fragrans namelyClerodendrumphilippinumSchauerhas antibacterial activity because it contains relatively high secondary metabolites [2].

Plant secondary metabolites can be obtained by extraction using solvent. Ethanol, ethyl acetate and *n*-hexane solvents have different dielectic constant (20° C) values, namely 22.40; 6.02 and 1.90 which causes the difference in the polarity of the three solvents. The ethanol, ethyl acetate and n-

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hexane solvents respectively are referred to as polar, polar and non-polar solvents. [3]. The choice of solvent for the extraction process is based on the solubility principle "like dissolve like". The nonpolar compounds dissolve only in nonpolar solvents and vice versa. The choice of solvent in the extraction process is an important factor[4-6]. The secondary metabolites found in *C. fragrans*are potential for *C. fragrans*as medicinal plants.

The Brine Shrimp Lethality Test (BSLT) is a method to determine the bioactivity of a compound from natural ingredients. Larvae *ArtemiasalinaL* is widely used for environmental studies, toxicity, and screening of bioactive compounds from plant extracts [7-8]. The BSLT test has a spectrum of pharmacological activity that is easy to perform, simple, fast, and does not require large costs with a 95% confidence level. The toxicity of compounds is expressed by the LC₅₀ value. The LC₅₀ value is an indication of the concentration of compounds that cause shrimp larvae death to 50% of the population. A sample is said to be toxic if it has a value of LC₅₀ <1000 µg/ml. The BSLT test can be used as a sedative, toxicity, insecticide test, and as a preliminary test for cytotoxic or anti-cancer compounds [9]. By using BSLT method, of 30 medicinal plants traditionally used in Bukota Tanzania, as many as 28 plants are safe to use with LC₅₀ valuesbetween 30-100 µg/mL [10].

Based on the description above, the authors were interested in testing the toxicity of *C. fragrans* leaf extracts which were extracted using ethanol, ethyl acetate and *n*-hexane as solvents. This study aims to determine the toxicity of secondary metabolites contained in *C. fragrans* leafextract as the basis to the development of *C. fragrans* plants as raw material herbal medicine.

2. Methods

2.1. Preparation of plant samples

The 4.1 kg of fresh leaves of *C. fragrans* were taken from the Simalungun area, North Sumatra. The leaves are separated from the stems, washed, drained, and dried in the room. The dry leaves were mechanically milled to obtain a crude leaf of powder *C. fragrans* as much as 520 g.

2.2. Plant extract preparation.

The 0.5 kg of *C.fragrans* leaf simplicia powder was extracted with *n*-hexane for 48 hours, then filtered and concentrated with a rotary evaporator. Part of the waste is macerated with ethyl acetate solvent for 48 hours, filtered and concentrated, obtained ethyl acetate extract concentrated. Part of the waste is extracted again with ethanol for 48 hours, filtered, and concentrated to obtain concentrated ethanol extract. The extraction process is carried out by maceration.

2.3. Toxicity brine shrimelethality test (BSLT)

The three types of *C. fragrans* leaf extracts were tested for their toxicity using the BSLT method with the following activity stages :

2.3.1.Culture of shrimp larvae. A total of 50.0 mg of eggs isArtemiasalinaLeach put into a hatchery filled with filtered seawater. After being aerated and irradiated with a 20 watt TL lamp for 24 hours, the eggs that have hatched into *nauplii*can be used as test animals.

2.3.2.Test sample preparation. As much as 40.0 mg of each extracted sample was dissolved in seawater to 20.0 mLin order to obtain a sample solution of 2000 μ g/mL. The samples of *n*-hexaneand ethyl acetate extracts which were rather difficult to dissolve in seawater. The extracts were added with 1% dimethyl sulfoxide (DMSO) of 1.0 μ L. A total of 2 mL of sample solution of 2000 μ g/mL was diluted to 20 mL in order to obtain a sample solution of 2000 μ g/mL. The 2 mL of sample solution of 200 μ g/mL was diluted to 20 mL and a sample solution of 20 μ g/mL concentration was obtained.

2.3.3. Test implementation. A total of 5.0 mLof the test sample from each concentration was put into a container that had diaries of 10 mL. Then, seawater was added to the container containing 10 shrimp

larvae that were 2 days old, and seawater was added to 10 mL, so that the final sample concentrations in each container were 1000, 100, and 10 μ g/mL. Each concentration was made three times (triple). The experiment container was placed under sufficient light for 24 hours.

2.3.4. Experimental data analysis. Observation of the number of Artemiasalina of dead from each sample concentration was carried out after 24 hours of the experiment. The mortality or mortality rate (%) was calculated by comparing the number of Artemiasalina that died with the total number of Artemiasalina tested. LC_{50} value is calculated by plugging the concentration and probit logs into the regression equation line. A substance is said to be active or toxic if the LC_{50} value is<1000 µg/mL. The LC_{50} value iscalculated by entering the log of concentration and probit into the regression equation line [8].

3. Results and discussion

The toxicity test results of leaf extract *C. fragrans*using the BSLT method are presented in Table 1. The results of the toxicity test showed that giving ethanol extract caused the highest mortality rate of the tested animals (100%), followed by ethyl acetate extract (96.67%) and *n*-hexane extract (93.30%). (Table 1).

Table 1. Toxicity test results of leaf extract C. fragrans with BSLT method

| No | Sample Test | Extract Concentration (µg/mL) | Number of Dead Larvae | | | Mortality (%) | LC ₅₀ (µg/mL) | |
|----|-----------------------|-------------------------------------|-----------------------|----|-----|------------------|-----------------------------|-------|
| | | | Ι | II | III | Total | _ | |
| 1 | Extract of <i>n</i> - | 10 | 2 | 2 | 3 | 7 | 23.33 | 41.97 |
| | hexane leaves | 100 | 8 | 9 | 9 | 26 | 86.67 | |
| | C fragrans | 1000 | 9 | 9 | 10 | 28 | 93.30 | |
| | | Control | 0 | 0 | 0 | 0 | 0 | |
| 2 | Extract of Ethyl | 10 | 2 | 3 | 3 | 8 | 26.67 | 37.50 |
| | Acetate leaves | 100 | 9 | 8 | 9 | 26 | 86.67 | |
| | C fragrans | 1000 | 9 | 10 | 10 | 29 | 96.67 | |
| | | control | 0 | 0 | 0 | 0 | 0 | |
| 3 | Extract of | 10 | 3 | 3 | 4 | 10 | 30.00 | 26.25 |
| | Ethanolleaves C | 100 | 8 | 9 | 9 | 26 | 86.67 | |
| | fragrans | 1000 | 10 | 10 | 10 | 30 | 100.00 | |
| _ | | control | 0 | 0 | 0 | 0 | 0 | |

Note: The total number of tested animals are 30 animals.

The high mortality rate causes the LC_{50} value of the ethanol extract to be the lowest, namely 26.25 $\mu g/mL$. This is probably because ethanol as a polar solvent easily attracts the active compounds contained in the sample, such as phenolic compounds, alkaloids, and terpenoids, were toxic to the zological system. Secondary metabolites flavonoids, triterpenoids, alkaloids, quinones and saponins are found in the ethanol extract of *C. fragrans*.

Plant extracts that had an LC₅₀ value of less than 20 µg/mL showed anticancer properties[9]. The BSLT test results of the ethanol extract of rantihitamfruit (*S. blumei*) which have an LC₅₀ value of 21.10 µg/mL [12] have an anticancer activity of leukemia cells L₁₂₁₀ [13] and contain the alkaloid steroid glycoside β 2-solanine compound [14]. The BSLT test results of methanol/water extract (70/30) from another plant of the genus Clerodendrum in Kenya, namely *Clerodendrummyricoides*, had lower mortality than *C. fragrans*. At a sample of *C. myricoides*10 µg/mL had a 16% mortality, at 1000 µg/mL it had a 76% mortality. The LC₅₀ value of *C. myricoides* was 204.66 <1000 (µg/mL) and the plant

was potentially bioactive [15]. Meanwhile, oral administration of *C. capitatum* extract to Wistar rats did not show hematological and biochemical side effects with an LD50 of 5g/Kg BW[16].

A substance is said to be active or toxic when the LC_{50} value<1000 (µg/mL).Based on this, the three *C. fragrans* leaf extracts were toxic and potentially bioactive with LC_{50} values of 26.25 µg/mL (ethanol extract), 37.50 µg/mL (ethyl acetate extract) and 41.97 µg/mL (*n*-hexane extract).The LC_{50} values of the three types of leaf extracts *C. fragrans* were in the range less than 1000 µg/mL, so the three extracts had the potential to be bioactive.

4. Conclusion

The leaf extracts *C* fragransare toxic and potentially bioactive with LC₅₀ values of 26.25 µg/mL (ethanol extract), 37.50 µg/mL (ethyl acetate extract) and 41.97 µg/mL (*n*-hexane). The extracts of *C*. fragranshas the highest toxicity with an LC₅₀ valueof 26.25 µg/mL compared to the ethyl acetate extract (LC₅₀ 37.50 µg/mL) and the extract *n*-hexane (LC₅₀ 41.97 µg/mL). The LC₅₀ values of the three types of leaf extracts *C*. fragranswere in the range less than 1000 µg/mL, so the three extracts had the potential to be bioactive. Leaf extract *C* fragrans have the potential to be developed as herbal medicine and it is necessary to research the isolation of bioactive compounds from leaf extract *C*. fragrans to develop the potential of Indonesian medicinal plants.

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