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ESTIMATING OF RAPD MARKER ASSOCIATED TO COLOR GENE IN Zinnia Elegans JACQ

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ABSTRACT

Flower color contributes mainly to the market value of ornamental plants including *Zinnia elegans* Jacq. The coloration of flowers is determined by several genes. The previous study showed that *Z*innia 16 loration is controlled by two independent genes, which are white color as dominant (chromogen) and its suppressor. The aim of this study is to 14 estigate marker that associated with chromogen based on RAPD. DNA from each of 240 in 18 iduals in M2 population was extracted from leaf by CTAB method (Doy 3 and Doyle, 1990). Fifteen primers were used to amplify DNA fragments. The PCR program was consisted of preheating at 94°C for 1 min, followed by 45 cycles of d 3 attraction at 94°C for 30 sec, annealing at 38°C for 30 sec, and elongation at 72°C for 1 min 30 sec. The last cycle was followed by a final extension cycle at 72°C for 7 min. Six among fifteen primers were selected and produced 60 loci which is 100% was polymorphic. The binary data were analyzed using Fisher Exact Test and Kendall Tau correlation in the SAS program to get correlation between markers and those traits. The result showed OPB17₁₄₀₀ and OPA18₁₅₀₀ markers were associated with chromogen in Zinnia.

Keywords: Zinnia elegans Jacq, flower color gene, association, primer, RAPD.

INTRODUCTION

Flower color is one of the most important traits in ornamental plants, dictating consumer interest and attraction. As such, it is a critica 4 actor for the commercial success of plants on the market. The inheritance of color in flowers is a large field, especially, as it varies in different species. Color can be dominant, recessive, or additive in nature, in different species, and in some plant species it can be a combination of these factors. Also some plants have two or more different genetic defects to make them white or 1 n-colored.

In general, a DNA marker linked to specific gene was developed by using near-isogenic lines (NILs), and their genetic linkage map at the specific gene (Y 2 mg et al., 1988; Martin et al., 1991; Paran et al., 1991). Various markers have been used to associate with different traits of crop plants. helmore et al. (1991) used bulk segregant analy, 2 technique to identify or link markers with various traits in many plants. DNA markers have been linked to fruit skin color in pear (Inoue et al., 2006), apple (XiaoWei et al., 2009) and grapes (Ren et al., 2000). Markers have been found to be associated with resistance for scab (Sestras et al., 2009) and columnar growth habit in apple (Costa et al., 2001). RAPD markers linked to sex were identified in pistachio (Kafkas et al., 2001) and guggal (Samantaray et al., 2010). QTLs were identified for grain yield in rice (Venuprasad et al., 2009), pod and kernel traits in peanut (Selvaraj et al., 2009) and rust resistance in eucalyptus (Zamprogno et al., 228). Markers for dwarfing in pear (YanLi et al., 2007), early flowering in eucalyptus (Domingues et al., 2006), pollen sterility in peach (Jun et al., 2004), precocity in walnuts (KeQiang et al., 2002) and seedlessness in grape (Mejia and Hinrichsen, 2003) were also identified and reported.

Kendall's tau value is defined as (concordant discordant) (concordant + discordant) values (Kendall, 1970). This value will be high (close to one) in the case of a positive relationship, positive meaning that both values are increasing in the same direction. Because it only involves the relative orderings of similarity values, it is relatively insensitive to 'outliers'. The aim of this study is to investigate marker that associated with chromogen based on RAPD using Fisher Exact Test and Kendall Tau correlation.

MATERIALS AND METHODS

Plant material 10 nd DNA extraction Total DNA was isolated by a modified CTAB method (Døyle and Doyle, 1987). DNA taken from each of 240 individuals in M2 population was extracted from 17. The primers used for PCR amplification are OPA 18, OPB 01, OPB 07, OF 3 10, OPB 17 and OPB 20. The PCR included preheating at 94°C for 1 min, followed by 45 cycles of denaturation 94°C for 30 sec, annealing at 38°C for 30 sec, an 8 longation at 72°C for 1 min 30 sec. The last cycle w1s followed by a final extension cycle at 72°C for 7 min. Random 10-base primers (Operon technology Inc.) were independently used for the polymerase chain reaction of RAPD analysis. Polymerase chain 9 actions were conducted in a volume of 10 μl containing 5 μl PCR mix Go Taq® Green (Promega), 0.25 μl 100 μM primer (Sigma-Proligo), 2.25 µl DN 1 genome as a template and 2.5 µl nuclease free water. The amplification products were analyzed by electrophoresis in 1.5% agarose gels (NuSieve 3:1 agarose). The gels were stained with ethidium bromide, visualized under 12 aviolet light and documented by camera. The RAPD bands were scored 1 as present or 0 as absent. The amplified fragments were

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evaluated using Fisher Exact Test and Kendall Tau correlation in SAS program to get correlation between markers and coloration phenotypes.

RESULT AND DISCUSSIONS

Two color phenotypes (colored phenotypic and colorless phenotypic) were observed in populations M_2 . There were 205 plants with colored flower and 35 plants with colorless (white). (Figure-1).



Figure-1. Two kinds of flower phenotype in Zinnia: coloured type (a,b,c); colourless type (d).

To identify specific RAPD markers associated with flower color phenotypes, 15 random primers were screened. Six primers showed polymorphism bands. Those

primers could amplified 60 bands which had molecular weight between 200 and 1900 bp (data not shown) (Figure-2).



Figure-2. Agarose gel electrophoresis of amplification products generated by primer OPB 07 (a) and OPB 17 (b).

The markers present in one of the bulks and absent in the corresponding bulk of contrasting phenotype were considered as polymorphic and identified as a putative marker for that flower coloration. Based on the Fisher Exact test and Kendall tau correlation, the primer

OPA18, OPB17 and OPB 07 have association with the coloration of Zinnia flower. OPA18, and OPB17 were associated with colorless (white) and OPB07 was associated with colored. (Table-1).

Table-1. A RAPD primer has associated with coloration flower in M2/population (Fisher Exact Test, 5% dan Kendall Tau correlation).

No.	Primers	Fisher exact test	Kendall Tau correlation
	OPA18-1500	0.0120	-0.1917
2	OPB17- 1400	0.0000	-0.3716
3	OPB07-200	0.0233	0.1560

This result supports a model involving two major genes proposed by Boyle and Stimart (1988). Presence of the anthocianidins pelargonidin and cyanidin is controlled by single dominant gene (An1). Carotenoid expression is conditioned by a recessive gene (ca) governing its presence and other genes controlling the distribution of carotenoid in ligules.

RAPD marker linked with colored phenotype (OPB 07-200) and two RAPD markers (OPB17-1400 and OPB18-1500) assosiated with colorless phenotype were obtained from M2 popula 11. Following the hypothesis of Boyle and Stimart (1988) this marker should be linked to a major gene on C locus (called modifier) that has the dominant effect on suppressing the colored flower. Those

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phenotypes were exhibited when the modifier gene worked incompletely.

We found that two RAPD markers (OPB181) and OPB17-1400) could be used to select colorless with probability a 1 high as approximately 19% and 37%, respectively. The marker was apparently useful for the putative markers individuals in 41 reeding program study of coloration in Zinnia flower. This is a first report on developing a DNA marker associated to the flower coloration phenotype in Zinnia. This experiment suggests the possibility of utilizing the DNA marker even in plant species, which have a highly heterozygous genome, without requiring a genetic linkage map and any DNA sequence information. Therefore, it seems to be an effective technique for conveniently developing selection markers in coloration of flower in Zinnia. In conclusion, we are now found that OPB18-1500 and OPB17-1400 associated with colorless.

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