ANTIOXIDANT, TOXICITY AND ANTIBACTERIAL PROPERTIES OF Ompu-ompu (Crinum asiaticum-L) ETHANOL EXTRACT

by Duma Riris

Submission date: 11-Mar-2019 03:13PM (UTC+0500)

Submission ID: 1091319611

File name: 3._476_pdf.pdf (388.13K)

Word count: 3488

Character count: 17322





3 ol. 11 | No. 3 | 1229 - 1235 | July - September | 2018 ISSN: 0974-1496 | e-ISSN: 0976-0083 | CODEN: RJCABP http://www.rasayanjournal.com

http://www.rasayanjournal.co.in

ANTIOXIDANT, TOXICITY AND ANTIBACTERIAL PROPERTIES OF Ompu-ompu (Crinum asiaticum-L) ETHANOL EXTRACT

Ida Duma Riris^{1,*}, Murniaty Simorangkir¹ and Albinus Silalahi¹

¹Chemistry Education Department, State University of Medan, North Sumatra, Indonesia.

*E-mail:dumariris@gmail.com

ABSTRACT

The Crinum plant varieties found, called "Ompu-ompu", identified as a type of Crinum asiaticum-L which used as a drug. The antioxidant, toxicity and antibacterial of the Crinum leaves and bulbs extracted with ethanol and treated by DPPH (2-2 diphenyl 1-1-picrylhydrazyl) method, Brine Shrimp Lethality Test (BSLT) method, and diffusion method respectively. It was found that both the leaves and bulbs have an antioxidant site with 54.21 ppm and 33.79 ppm respectively through IC₅₀ test 25.8 ppm which used vit.C as a control. Furthermore, it was found that their toxicity measured by LD₅₀ is 243.331 and 507.838 respectively. In addition, the antibacterial test showed that the widest inhibition of the leaves and bulbs are 15.7 mm and 12.4 mm against Escherichia coli respectively and 12.3 mm and 11.5 mm against Staphylococcus aureus. Phytochemical screening indicated the presence of alkaloids, flavonoid, saponins and tannins.

Keyword: antibacterial, antioxidant, Crinum Asiaticum L, toxicity, phytochemical

© RASĀYAN. All rights reserved

INTRODUCTION

The *Crinum* is a flowering plant, which is one of the large genus in the Amaryllidaceae family. There are 180 species of this plant, which are spread out in Asia, Australia, Africa and America, which have secondary metabolite content especially alkaloids¹. It is typically white to pink and grows in tropical and sub-tropical regions. One type of the plant is found in *Haunatas* Village, Tobasa District, North Sumatra Indonesia, with typically white and its height reaches one and a half meters. Local people call it "*Ompu-Ompu*" as shown in Fig.-1.



Fig.-1: "Ompu-ompu"- Crinum asiaticum L.

The plant is classified into a *Crinum asiaticum-L* species of Genus Amaryllidaceae family based on the family identification, which identified in Pharmacy laboratory of North Sumatra University. Generally, the local community uses it as a medicine.

Rasayan J. Chem., 11(3), 1229-1235(2018) http://dx.doi.org/10.31788/RJC.2018.1133090



The leaves have a function to overcome the swelling of the human body due to impact or sprains. It is carried out applying oil on to the leaves, and then heat and place it on the swollen parts. Phyto screening results of bulbs of *Crinum asiaticum* L. var. sinicum BAKER from China has found 21 compounds that have activity as anti-tumor².

The investigation of *Crinum aurebensaiton* through alkaloid isolation process³ revealed that it contains dehidro anhidroclorine. Likewise, research on the *Crinum jagus* and *Crinum glaussum* species found that alkaloids from extracts of bulbs showed inhibition of acetyl colinesferase, and it results in the depressed levels of acetylcholine in the brain associated with Alzheimer's disease⁴. It reported that the alkaloid compounds have protective properties due to the presence of lycorine compounds in the leaves which are considered as an important source of natural antioxidants^{5,6}.

It has been identified that *Crinum* species contains 118 different alkaloids⁷ which was determined through Phytochemical test⁸. The alkaloids are generally beneficial to human health such as in weight loss program, antinociception, antihitperalgesic, sedative, and antioxidant. Furthermore, it can reduce the blood sugar levels or it can be used as an antidiabetic agent^{9,10}. In other sources, also reported that it can be used as an anti-tumor, immunostimulan, analgesic, and anti-bacterial ¹¹. It is considered as an antioxidant because it is not only can stabilize free radicals but also can inhibit the formation of the free radicals which cause oxidative stress^{12,13}. As previous researchers tested the antioxidant by dpph method at 517 nm¹⁴⁻¹⁷, in this study for antioxidants performed with a spectrophotometer with 517 nm.

The antioxidants can be found in several types of plants, and it reported that those derived from plants are better than synthetic antioxidants, because the synthetic one can negatively affect human health when consumed¹⁸. The other important point of concerns in utilizing the plant which is toxic properties. The toxicity of a plant is considered as an indication of the potential of compounds as a medicinal plant (bioactivity)^{19,20}. There are 7 types of alkaloids from cognates with *Crinum* that have significant toxicity and activity against antibacterial and cancer resistance²¹ which were tested by the Brine Shrimp Lethality Test (BSLT)²².

The investigation of the existing *Crinum* plant at various sites expands the crop inventory because the production of an organic compound secondary metabolite of a plant is closely related to environmental factors where it grows²³. Investigation results of the *Crinum* from the *Haunatas* Village showed that the antioxidant, toxicity, and antibacterial of the plant are highly valuable and presented in detail in this paper.

EXPERIMENTAL

Collection of Plant

Crinum asiaticum L is obtained from the peasant Haunatas Village North Sumatera Indonesia.

Preparation of Plant Extract

A total of 15 kg freshly *Crinum asiaticum-L* has separated the leaves and bulbs, and then washed with 500 ml of distilled water and sliced with a knife. Before drifting, the blanching process was done by boiling for 5 minutes used 0.5% citric acid at temperature $100^{0}C^{24}$, air dried at room temperature for 14 days. The dried leaves and bulbs blendered mashed to get the powder extract. The next, 500 gr of leaves and bulbs powder were macerated with 2 liters of ethanol sol to 3 x 24 hours. The same process was carried out 3 times then the results were filtered with Buchner. The filtrate was concentrated by vacuum rotary evaporator until the ethanol extract is collected. Further bioactivity measurement was carried out by phytochemicals, antioxidants, toxicity, and antibacterial tests.

Phytochemical Screening

The Chemical screening of Ethanol extract plant was found by the Harborne method, 19878.

Antioxidant Assav

The DPPH (2-2 diphenyl 1-1- picrylhydrazy) method is used for the antioxidant test by making ascorbat acid with concentration 5, 10, 25, 50, dan 100 ppm as a control. Furthermore, DPPH powder is dissolved in methanol p.a and made a comparative test solution of 2 mL inserted into the test tube by adding the solution of 2 mL DPPH 0.1 mM, and stirred with vortex until homogeneous. The homogeneous solutions

Vol. 11 | No. 3 | 1229 - 1235 | July - September | 2018

were incubated in the dark for 30 minutes, and measured uptake at a 517 nm using the visible spectrophotometer. Based on this, the percentage of inhibition is calculated by the formula,

Activity (%) =
$$[(A-B)/A] \times 100 \%$$
 (1)

Where, A is the absorbance of control (DPPH solution without the sample), B is the absorbance of the DPPH solution in the presence of the sample (extract/ ascorbic acid)²⁵.

The Inhibition Concetration $50(IC_{50})$ value is the concentration of antioxidants (ppm) which is able to inhibit 50% of the free radicals, whose value is indicated by the intersection of the intermediate line of 50% inhibition power with the concentration axis, then incorporated into the equation,

$$Y = a + bX \tag{2}$$

Where Y = 50, and the X value showed IC₅₀.

Toxicity Test

The toxicity test used Brine Shrimp Lethality Test (BSLT). The fifteen vials were prepared with five different concentrations of test solution such as 1000, 500, 200, 100, 10 ppm. The analysis was done for three-time replication include the control. The main solution was prepared by dissolving 0.2 g of extract into seawater extract until 100 ml of volumetric flask and homogenized. The main solution was taken 2500, 1250, 500, 250, 25 ppm respectively to make a solution with concentrations of 1000, 500, 200, 100, 10 ppm. The solution was stirred until homogeneous and evaporated until dry. Each concentration was made in 3 vials, then into each vial inserted 5 ml of seawater and 10 naupli for 2 hours then observed the number of death shrimp larvae with a magnifying glass.

Furthermore, the LC_{50} was calculated with the concentration of test substance that can kill shrimp larvae as much as 50% with SPSS 20 application 15 h this case, the sample was very to the shrimp larvae Artemiasalina Leach if the value of LC_{50} <30 μ g / ml, toxic if 30 μ g / ml \geq LC_{50} <1000 μ g / ml, and not toxic when LC_{50} > 1000 μ g / ml²⁶.

Antibacterial Screening

Instruments sterilization were important before test analysis. Glasses were sterilized in the oven with 100°C for 2 hours. On the other hand, met were sterilized with a spray-mounted light, and for heat-resistant appliances and the high-temperature medium was sterilized in an autoclave at 121oC, 2 atm for 15 minutes. 2.3 g of NA (Nutrient Agar) were added and mixed with 100 mL aquabidest in the beaker glass until homogenous. This homogenous mixture was autoclaved for sterilization. The UV light was ignited on the laminary for 15 minutes and blown for 10 minutes, then poured into the petri dish as a medium.

Escherichia coli as a sample were reconstructed before being used for antibagierial tests. The bacteria were cultured on a bend sterilized agar, then incubated for thours at 37 ° C. The Bacterial suspension was spread using a sterile cotton swab on nutrient agar plat. The turbidity of each bacterial culture was adjusted to 0,5 McFarland Standard then incubated for 24 hours^{27,28}. The ethanol extract was dissolved in concentrations of 0%, 2.5%, and 5%.. Chloramphenicol 0.02% were used as reference drug for bacteria (as much as 0.002 grams of chloramphenicol dissolved in 10 mL of distilled water (0.025) (w/v)²⁹. Zone of Inhibition was measured in mL.

RESULTS AND DISCUSSION

Based on the fitoscreening test of the leaves and bulbs ethanol extract (Table-1). It was found that the leaves contain an alkaloid, flavonoid, saponin, and tannin, as well as in the bulbs except for tannin. This suggests that the "Ompu-ompu" may have a variety of bioactivity such as antibacterial, antioxidant, toxicity, anticancer and others³⁰⁻³¹.

The antioxidant test for the ethanol extract is used with the DPPH method which used UV-Vis at 517 nm. The result showed that the leaves and bulb shave the antioxidant value of 54.21 ppm and 33.79 ppm respectively with the 25.8 ppm of vit. C as the control solution (Table-2). These indicated that the "Ompuompu" has antioxidant properties, which presented in Fig.-2. This is similar to the antioxidant activity of methanol extracts of crops found in Bangladesh which were measured by DPPH (1,1-diphenyl-2-picrylhydrazyl) method, with the vitamin C as a control ¹² and also found at the Cauvery river basin, Thanjavur, Tamilnadu India⁷. Isolation and identification results showed the phenolic compounds as the

previous research³³. This phenolic concentration indicates the presence of antioxidant activity³⁴. The toxicity of the crinum leaves and bulbs ethanol extracts graphically shown in Table-3 and Fig.-3.

Table-1: Phytochemicals of Crinum asiaticum L Ethanol Extract of Bulbs and Leaves

No.	Phytochemicals	Leaves	Bulbs
1.	Alcaloid	+	+
2.	Flavonoid	+	+
3.	Saponin	+	+
4.	Tanin	+	(#)

The death of naupli in hour difference is seen on the vertical axis, and the leaves and bulbs extract concentration ($\mu g / mL$) is visible on the horizontal axis. By using the SPSS version 22, It was found that the LC₅₀value of the leaves and bulbs are 243,331 ppm $\frac{1}{8}$ 507,838 ppm respectively. It shows that both the leaves and the bulbs are showing toxic properties. The result is in line with the previous research¹⁹ which shows that the toxic properties of a substance are known if the value of LC₅₀is <1000 ppm for the extract and ≤ 30 ppm for a compound. It is presumed that the "Ompu-ompu" is considered as a potential and promising medicinal plant. Because the plants have toxic properties ^{18,23} which are similar to the toxicity of the 7 alkaloids found in Licorisaurea(Amaryllidaceae) that have a significant antibacterial activity and even potentially as an anticancer²⁰.

Table-2: Antioxidant of Crinum asiaticum L Ethanol Extract

Extract	Concentration (ppm)	Absorbance	% Resistance	IC ₅₀ (ppm)
	5	0.233	57.48	
	10	0.2271	58.55	
Bulbs	25	0.1666	69.59	
	50	0.1382	74.78	33.79
	100	0.095	82.26	
	5	0.2496	54.45	
	10	0.1805	67.06	
Leaves	25	0.1125	79.47	
	50	0.092	83.21	54.21
	100	0.0824	84.96	
	5	0.3209	41.44	
	10	0.1693	69.11	
itamin C	25	0.1406	74.34	
	50	0.1243	77.32	25.08
	100	0.0912	83.36	

The antibacterial agent of the leaves and 10 lbs ethanol extracts at various concentrations of 2.5% and 5% was indicated by its resistance diameters against *Escherichia coli* and *Staphylococcus aureus* (Table-4). The extent of inhibition obtained for the *Escherichia coli* bacteria was 12.4 (the leaves) and 15.7 (the bulbs); and the *Staphylococcus aureus* with an area of 11.5 (the leaves) and 12.3 (the bulbs).

Tabel-3: Toxicity of the '	"Ompu-ompu" - Crin	um asiaticum L.I eaves at	d Bulbs Ethanol Extract

Extract ethanol	Concentration (ppm)	Mortality	% Lethality	Total (n)	LC ₅₀ (ppm)
	10	2	20	10	154.546
	100	5	50	10	243.349
Bulbs	200	7	70	10	300.169
	500	9	90	10	507.838
	1000	10	100	10	Infinity
	10	3	30	10	52.435
	100	4	40	10	98.565
Leaves	200	6	60	10	130.495
	500	8	80	10	243.331
	1000	10	100	10	29985.832

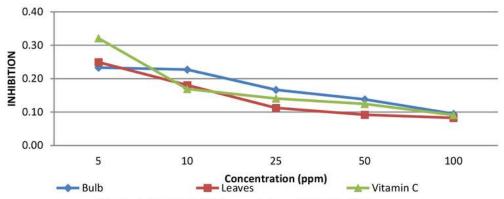


Fig.-2: Antioxidant of Crinum asiaticum L Extract Graph

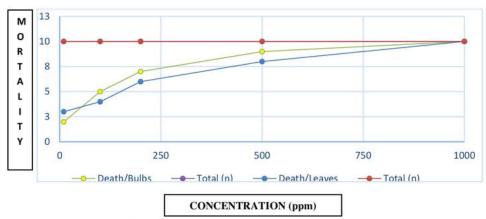


Fig.-3: Toxicity Graph of Crinum asiaticum L' Bulbs and Leaves

The inhibition power of the bulbs ethanol extract is greater than the leaves ethanol extracts against both bacteria. And when compared to the chloramphenicol controlled compound, the resistance power of the leaves and bulbs ethanol extract to the *Escherichia coli* was 34.73-49.06%; and for the *staphylococcus aureus* was 35.04% - 49.56%. It means that the antibacterial power of the Crinum found as strong as the previous findings ²⁹, such as the antibacterial species of *Boophone disticha (Amaryllidaceae)*, which has been processed and used as a traditional medicines ²².

Table-4: The Antibacterial activity of C.asiaticum L	L Bulb and Leaves of Extract Ethan	ol
--	------------------------------------	----

Extract Ethanol		Antibacterial Bacterial Activity			Cloramphenicol	
	D	The diameter of				
of Plant	Bacteria	The concentration of Extract (%)			(0,02%)	
		0	2,5	5		
	Escherichia coli	0	11,8	15,7	32,7	
Bulbs	Staphylococcus aureus	0	7,1	12,3	35,1	
	Escherichia coli	0	8,2	12,4	35,7	
Leaves	Staphylococcus aureus	0	6,8	11,5	23,2	

CONCLUSION

It is concluded, that the "Ompu-ompu"- Crinum asiaticum-L which located in the Haunatas Village contains alkaloid compounds which have strong antioxidant, toxicity, and antibacterial properties. It is highly recommended that the crinum plant is considered as a potential herbal medicinal plant and endorsed to carry out in detail research to trace other potential activities.

ACKNOWLEDGMENT

I would like to thank the State University of Medan, who funded this research with a contract No. 206 A/UN 33.8/PL/2017

REFERENCES

- 1. J. R. Lewis, ChemInform., 33(26), (2010), DOI: 10.1002/chin.200226247
- Q. Sun, Y. -H. Shen, J. -M. Tian, J. Tang, J. Su, R. -H. Liu, W. D. Zhang, Chemistry and Biodiversity, 6(10), 1751(2009), DOI:10.1002/cbdv.200800273
- 3. C. G. Guerrieri, N. B. Pigni, J. P. de Andrade, V. D. dos Santos, F. Binns, W. de Souza Borges, J. Bastida, *Arabian Journal of Chemistry.*, **9(5)**, 688(2016), **DOI:**10.1016/j.arabjc.2015.07.009
- P. J. Houghton, J. M. Agbedahunsi and A. Adegbulugbe, *Phytochemistry.*, 65(21), 2893(2004), DOI:10.1016/j.phytochem.2004.08.052
- 5. S. Ilavenil, B. Kaleeswaran, P. Sumitha, D. Tamilvendan and S. Ravikumar, *Saudi., Journal of Biological Sciences.*, **18(2)**, 181(2011), **DOI:**10.1016/j.sjbs.2010.11.001
- P. Cao, D. -S. Pan, S. Han, C. -Y. Yu, Q. -J. Zhao, Y. Song and Y. Liang, Archives of Pharmacal Research., 36(8), 927(2013), DOI:10.1007/s12272-013-0089-3
- N. Thi Ngoc Tram, T. Titorenkova, V. St. Bankova, N. Handjieva and S. Popov, Fitoterapia, 73(3), 183(2002), DOI:10.1016/s0367-326x(02)00068-0
- J. B. Harborne, *Phytochemical Methods*, Chapman & Hall London International Diabetes Federation, Atlas Sixth Ed., 2013 (www.idf.org/diabetesatlas).
- Y. J. Jeong, E. –H. Sohn, Y. –H. Jung, W. –J. Yoon, Y. M. Cho, I. Kim, S. C. Kang, Biomedicine & Pharmacotherapy., 82, 35(2016), DOI:10.1016/j.biopha.2016.04.067
- W. D. Ratnasooriya, S. A. Deraniyagala, S. D. N. K. Bathige and H. D. I. Hettiarachchi, *Journal of Ethnopharmacology.*, 97(1), 123(2005), DOI:10.1016/j.jep.2004.10.024

- 11. M. A. Rahman, R. Sharmin, M. N. Uddin, M. S. Rana and N. U. Ahmed, Bangladesh Journal of Microbiology., 28(1), (2012), DOI:10.3329/bjm.v28i1.11801
- TOPIC 07- Oxydative stress and aging. Archives of Cardiovascular Diseases Supplements., 4, 18(2012), DOI:10.1016/s1878-6480(12)70793-0
- V. Lobo, A. Patil, A. Phatak and N. Chandra, *Pharmacognosy Reviews.*, 4(8), 118(2010), DOI:10.4103/0973-7847.70902
- 14. J. Srivastava, D. Desh, P. Poonam, Rasayan Journal Chemistry., 8(2), 161(2015).
- J. A. O. Olugbuyiro, S. B. Anuoluwapo, O. A. Alaba, S. T. Olugbenga, A. Oyeronke, *Rasayan Journal Chemistry.*, 11(2), 798(2018), DOI:10.31788/rjc.2018.1121823
- 16. Y. Taufik, W. Tantan, G. Yudi, Rasayan Journal Chemistry., 9(4), 762(2016).
- 17. S. L. Deore, S. S. Khadabadi, S. P. D. Patel, M. S. Jaju, N. R. Junghare, T. P. Wane, R. G. Jain, Rasayan Journal Chemistry., 2(1), 129 (2009).
- 18. S. Azam, M. Mahmud, M. Naquib, S. Hossain, M. Alam, M. Uddin, M. Hasan, *Biomedicines.*, 4(3), 17(2016), DOI:10.3390/biomedicines4030017
- T. J. Kim, J. L. Silva, M. K. Kim and Y. S. Jung, Food Chemistry., 118(3), 740(2010), DOI:10.1016/j.foodchem.2009.05.060
- 20. A. Azhari, Scientiae Educatia., 3(2), (2014), DOI:10.24235/sc.edu.v3i2.539
- 21. J.-H Song, L. Zhang, L. Y. Song, Fitoterapia., 95, 121(2014), DOI:10.1016/j.fitote.2014.03.006
- 22. B. Meyer, N. Ferrigni, J. Putnam, L. Jacobsen, D. Nichols and J. McLaughlin., *Planta Medica.*, 45(5), 31(1982), DOI:10.1055/s-2007-971236
- 23. T. M. Kutchan, Plant Physiology., 125(1), 158(2001), DOI:10.1104/pp.125.1.58
- 24. I. Doymaz, Food and Bioproduct Processing., 88(2-3), 124(2010), DOI:10.1016/j.fbp.2009.09.003
- M. A. Rahman, R. Sharmin, M. N. Uddin, M. S. Rana and Ahmed., Bangladesh Journal of Microbiology., 28(1), (2012), DOI:10.3329/bjm.v28i1.11801
- 26. M. Steven and Russell Molyneux., CRC Press Boca Raton Ann Arbor London Tokyo. (1985)
- A. Zapata and S. Ramirez-Arcos, Current Microbiology., 70(6), 907(2015), DOI:10.1007/s00284-015-0801-2
- 28. J. H. Wicks, R. L. Nelson and G. E. Krejcarek, Journal of Clinical Microbiology, 17(6), (1983)
- 29. M. Steven and R. Molyneux, Bioactive Natural Products. CRC Press, Tokyo, 3-14 (1993).
- D. W. Lambe and W. Wayne Laslie, Anaerobic Bacteria., 51–60,(1980), DOI:10.1007/978-1-4613-3159-9
- S. A. Adesanya, T.A. Olugbade, O. O. Odebiyl and J. A. Aladesanmi, *Pharmaceutical Biology.*, 30(4), 303(1992), DOI:10.3109/13880209209054019
- L. Cheesman, J. J. Nair and J. Van Staden, *Journal of Ethnopharmacology.*, 140(2), 405(2012), DOI:10.1016/j.jep.2012.01.037
- 33. M. S. Blois, Nature., 18, 4617(1958), DOI:10.1038/1811199a0
- Q. Sun, W. D. Zhang, Y. H. Shen, C. Zhang and H. L. Li, *Chinese Chemical Letters.*, 19(4),447(2008), DOI:10.1016/j.cclet.2008.01.022
- C. N. Kunyanga, J.K. Imungi, M. W. Okoth, H. K. Biesalski and V. Vadivel, LWT Food Science and Technology., 45(2), 269(2012), DOI:10.1016/j.lwt.2011.08.006

[RJC-3090/2018]

ANTIOXIDANT, TOXICITY AND ANTIBACTERIAL PROPERTIES OF Ompu-ompu (Crinum asiaticum-L) ETHANOL EXTRACT

EXTRAC	T				
ORIGINALITY RE	PORT				
% SIMILARITY IN	IDEX	5% INTERNET SOURCES	6% PUBLICATIONS	2% STUDENT PA	\PERS
PRIMARY SOUR	CES				
	w.apponet Source	connect.in			2%
Jar G. I ant hex Ple Agr	nes S. Reyes ibacte kane e urotus	on R. Dulay, Lui Malasaga, Sofr , Ching T. Hou. rial activities of xtracts of Lentin s djamour", Bioc ral Biotechnolog	onio P. Kalaw "Antioxidant a acetonitrile an nus tigrinus ar atalysis and	r, Renato and nd	1%
.)	W.rasa net Source	ayanjournal.co.ir	1		1%
Sta		d to Universitas versity of Surab		oaya The	1%
D S	Sukma	wati, D Dellanei	ra, A Risandi		_1

D Sukmawati, D Dellanerra, A Risandi.
"Screening the capabilities of Indonesian indigenous mold in producing cellulase

<1%

enzyme", IOP Conference Series: Materials Science and Engineering, 2018

Publication

6	fr.slideshare.net Internet Source	<1%
7	rasayanjournal.co.in Internet Source	<1%
8	www.slideshare.net Internet Source	<1%
9	Heng Luo. "Antioxidant activities of edible lichen Ramalina conduplicans and its free radical-scavenging constituents", Mycoscience, 04/08/2010 Publication	<1%
10	www.science.gov Internet Source	<1%
11	Edison Osorio, Gabriel Jaime Arango, Nora Jiménez, Fernando Alzate et al. "Antiprotozoal and cytotoxic activities in vitro of Colombian Annonaceae", Journal of Ethnopharmacology, 2007 Publication	<1%
12	Qi, Hui Juan, Yu Yang, Li Bo Wang, Ya Qin Xu,	<1%

Qi, Hui Juan, Yu Yang, Li Bo Wang, Ya Qin Xu and Shu Ping Qu. "Determination of Bioactive Ingredients in Five Pumpkin Seeds Cultivars", Advanced Materials Research, 2012. banglajol.info
Internet Source

- <1%
- Simorangkir, Murniaty, Tonel Barus, Ribu Surbakti, and Partomuan Simanjuntak.
 "Isolation and Toxicity of Steroidal Alkaloid Glycoside from Fruits of Ranti Hitam (Solanum blumei Nees ex Blume)", Asian Journal of Chemistry, 2016.

<1%

Publication

Pekik Wiji Prasetyaningrum, Anton Bahtiar, Hayun Hayun. "Synthesis and Cytotoxicity Evaluation of Novel Asymmetrical Mono-Carbonyl Analogs of Curcumin (AMACs) against Vero, HeLa, and MCF7 Cell Lines", Scientia Pharmaceutica, 2018

<1%

Publication

Exclude quotes On

On

Exclude bibliography

Exclude matches

Off