RESEARCH ARTICLE

Fabrication of Silica-Based Chitosan BiocompositeMaterial From-from Volcanic Ash and Shrimp Husk by Sol Gel Method for AbsorbAdsorbent

LISNAWATY SIMATUPANG^{1,2}, MANIHAR SITUMORANG^{2*}, HARLEM MARPAUNG^{1*}AND RIKSON SIBURIAN¹

¹Postgraduate Study Program, Departmentof

Chemistry,FacultyofMathematicsandNaturalSciences UniversitasSumateraUtara,Jl. Bioteknologi, Kampus USU Padang Bulan,Medan,Sumatera Utara, 20155, INDONESIA ²Department of Chemistry, FacultyofMathematics andNatural Sciences, UniversitasNegeri Medan,Jl. WillemIskandarPsr. V, Medan,Sumatera Utara, 20221,INDONESIA

*Corresponding author : Manihar Situmorang, Department of Chemistry, Faculty of Mathematics and Natural Sciences, Universitas Negeri Medan, Jl. Willem Iskandar Psr. V, Medan, Sumatera Utara, 20221, INDONESIA, Phone: (062)-61-6613365, Faximile: (062)-61-6613319, E-mail: msitumorang@unimed.ac.id

Abstract

The utilization of volcanic ash and shrimp shell waste into composite material is very interesting to discussas a strategy to makenaturally available materials intovaluable, stable and low cost absorbadsorbent. The study is aimed to fabricatethesilica-based chitosanbiocompositematerial from volcanic ash of Sinabung mountain and shrimp husk by sol gel method to be used as an absorbadsorbent. The procedure wasperformed by converting volcanic ash into silica salt, extracting the chitosan from shrimp skeleton, and the synthesis of the silica-chitosanbiocomposite by mixing the silica with chitosan in the aid ofbifunctionalglutaraldehydecross-linking agent. The silica-based chitosancomposite was successfully prepared from the mixture of 20 mLNa₂SiO₃with 2% chitosanto obtain biocompositematerial (SChC1). A rapid and simple method to bind the silica with chitosan is achieved. An amorphoussmall crystalline composite (area $2\theta = 20-24^{\circ}$) is obtained. Absorption Adsorption capacity of the composite material has been evaluated, where it has the capability to bind heavy metal ion due to the presence of amine group in the material surface. The prepared material of silica-chitosanbiocomposite has the potential to be used as an absorbadsorbent.

Keywords:Sol-gel method, Silica-chitosan biocomposite, Volcanic ash, AbsorbAdsorbent, Heavy metal

Introduction

The use of silica-chitosan composite become promising material for its favour in the functional group that can be modified for different applications in analytical methods. Biocomposite is avaluable material forpractical application in Analytical Chemistry due to its

Commented [NN1]: For what adsorbate?

-1	Commented [NN2]: What is the concentration?
-	Commented [NN3]: %w/v?
-{	Commented [NN4]: What is the meaning?
-{	Commented [NN5]: Adsorption?
-(Commented [NN6]: Be specific!

biocompatibility and biodegradability, strong bio-adhesivity and film-forming ability, and excellent reusability¹.Various types of biocomposite materials have been developed andapplied for many purposes^{2–4}.The application of biocomposite for adsorbent materials to removeheavy metalshas been reported⁵.The biocompositematerial has also been applied for removing pollutant materials such as cation, anion, dyes, organic compounds, proteins, and heavy metals⁶.The fabrication of composite by utilizing naturally available materials is very challenging to obtain valuable, stable and low cost biocompositematerial forabsorbadsorbent. One of the abundant raw materials in North Sumatera is volcanic ash that can be utilized as a valuable silica-basedcomposite material.

The eruption of Mount Sinabungin Kabupaten Karo, North SumateraIndonesia 2013-2018, where many villages are covered with volcanic ash that demolished local plants, and the abundant of the ash becomes a problem for horticultural agriculture in the impact areas^{7–}⁼⁹. The locally available geological ash which has main content of silica (SiO₂)needs to be utilized to become a valuable material.

Some of the very potential properties of the silica such as acid resistance, very strong mechanical properties, large porosity, heat resistance, and resistance to microbial attack make it potential for the manufacture of low cost silica-based composite materials to be applied for many products^{10,11}.Previous study has showed that the synthesized silica based material obtainedfrom volcanic ash has successfully producedsilica material that has potential to be used as an adsorbent¹².However, its low surface ability to interact with the target ions required the silica to be modified with organic material such as chitosan to enable the composite material to become an effective adsorbent and other purposes^{13,14}.

Chitosan is a natural polycationic polysaccharide considered to be an important material to be combined with silica in the preparation of biocomposite material¹⁵⁼¹⁷.Naturallyavailable organic material ofshrimp shell waste can be used as the source of the chitosan. The chitosan with good biocompatibility, high adhesion to the surface, low toxicity and structural variability with amine functional group is a good substance to be used in many applications, including the<u>adsorb</u>ents^{18–20}. The presence of functional amine group in biomaterial is very important to be employed in various applications such as immobilization of enzyme^{21–24}, and therefore the chitosan with the amine group can be employed for many purposes, including thesensor and biosensor^{25–27}. The physical and chemical properties of chitosan with amino and hydroxyl make it compatible in the preparation of biodegradable silica-chitosan composite for various applications²⁸. Although the chitosan contains high

Commented [NN7]: Karo District?

amine and hydroxyl groups, it does not have specific selectivity for certain ions such as complex pollutants in water or wastewater^{29,30}. The chitosan without modification has low mechanical strength and low solubility in acidic medium. The amine group on chitosan in acid solution causes protonated chitosan. Therefore, modification is needed to optimise the compatibility of the chitosan for further applications.

Sol-gel process is assigned to be a good strategy in the preparation of silica-chitosan composite³¹.The sol-gel method has successfully been used for the composite systems of inorganic-organic biomaterials^{32,33}. The reaction is proceeded in the presence of polymeric organic molecule containing functional groups that can immobilised inorganic compounds.Preparation of silica-chitosan composite material has also been carried out in the presence of tetraethoxysilane as precursor of SiO₂³⁴.Formation of composite nanomaterial by sol-gel method with intention to modify the silica surfacehas also been been reported^{35,36}.The synthesis with this technique is easyto perform, the raw material is cheap, and the yield isrich withmultifunctional groups³⁷.Sol-gel method provides many advantages: the binding process taking place simultaneously, the reaction condition can be performed at room temperature, the procedure is simple, the end product is high in purity, homogeneous, uniform, and small in size¹³. This method is assigned to be effective in the preparation of silica-based biocomposite materials because the combinations of the hardness properties of the silica and the functional groupfromchitosan are utilized. The study is aimed to fabricatea silica-based chitosanbiocompositematerial fromraw material of volcanic ash of Sinabung mountain and shrimp husk skeletonby sol gel method. The characterization of the biocompositehas been performed, and the potential application of the composite as an adsorbent material to remove heavy metals was also demonstrated.

Commented [NN8]: All heavy metals?

Experimental Procedures

Chemical and Reagents

Reagent grades of HNO₃, NaOH, HCl, CH₃COOH, ethanol, CdCl₂, and glutaraldehyde25% were obtained from E-Merck and used without purification. All chemicals were dissolved inMilli-Q water. The volcanic ash containing silica was obtained from the eruption area of MountSinabung at Brastepu village, Kabanjahe, North Sumatera, Indonesia.

The chitosan was prepared from Lobster shrimp huskthat was collected from shrimp waste at Percut Sei Tuan, Deliserdang, North Sumatera, Indonesia.

Apparatus

Sample preparation was carried out by using an oven, furnace, sieve (200 mesh), magnetic stirrer, hot plate, ultrasonic (Braunsonic), desiccator, solid-phase extraction column, glass ware and plastic apparatus. The measurements was made by using a Fourier Transform Infra-Red (FTIR) Bruker spectrometer equipped with a Digitech detector (Shimadzu), X-Ray Fluorescence Spectrometer (XRF) (Perkin Elmer 3110), Rigaku ZSX X-Ray Diffraction (XRD) (Shimadzu XRD 6000), Atomic <u>AbsorptionAdsorption</u> Spectrophotometry (AAS) (Z-2000 series), Scanning Electron Microscopy Electron Dispersive X-ray (SEM-EDS) (Zeiss type EPOMH 10 Zss), Gas Sorption Analyser (GSA) with NOVA Quantachrome instruments version 2.2.

Procedures

Theprocedures in thefabrication of biocompositeare consisted of sample preparation of silica from volcanic ash,extraction of chitosan from shrimp skin, preparation of silica-based chitosan composite, and characterization of the compositefollowed the procedures explained earlier¹³. The experiment for testing of adsorbent capacity toward heavy metals is also carried out by using modifed procedures explained in the references^{38,39}.

Preparation of Silicate from Volcanic ash

The preparation of silicate from volcanic ash was carried out by soaking volcanic ash (20 g) into 120 mL 6M HCl for 24 hours, followed by dried the material in an oven (120° C for 6 hours), and the weight of the crude product was recorded. The crude was then destructed by using 156 mL 4M of NaOH at the furnace (500 °C and 30 minutes) until the viscous obtained, followed by addition of 200 mL water and let the solution to settle for 24 hours. The solution was then filtered to produce sodium silicate (Na₂SiO₃) as silicate raw material.

Extraction of Chitosan from Shrimp Husk.

Preparation of the chitosan from shrimp skin skeleton was conducted by cleaning the Lobster shrimp husk, followed by washing, drying, and grinding raw material shrimp shells into powder. Extraction of chitosan from the raw material was performed by using of a 3.5% NaOH solution with a ratio of 1:10 (w/v) at 65 °C for 2 hours to form chitosan crude, and the procedures are followed by the deproteinase and demineralization process by using of 1N

HCl solution with the ratio was kept at 1:15 (w/v) for 30 minutes troom temperature. The extracted chitosan was then reacted with 60% (w/v) NaOH solution at a ratio of 1:10 (w/v) at 125 °C for 2 hours. The mixture solution was then cooled, filtered, washed to neutral pH condition, and left to dry under sunlight. The chitosan obtained from this procedure was then used as a raw material for the preparation of silica-chitosan biocomposite.

Fabrication of Silica-based Chitosan Biocomposite

The fabrication of silica-based chitosan biocomposite was performed by mixing of sodium silicate with chitosan in a plastic container, and the synthesis was completed by addition of glutaraldehyde crosslinking agent. Various concentration of chitosan solution was prepared by successive addition of (2, 3,4, and 5) g chitosan into flask containing of 100 mL of acetic acid (2% v/v), and the mixture was then stirred for 1 hour to form (2%, 3%, 4%) and 5%) chitosan solutions, followed by addition of 1 mL glutaraldehyde solution (5%) and stirred vigorously for 5 minutes. A mixture of chitosan and glutaraldehyde solution is then poured into a 20 mL sodium silicate solution under constant stirring in a magnetic stirrer, followed by neutralization by drops addition of 3M HCl until the was pH 7.0. The immobilization proceeded overnight to form silica-chitosan gel. The gel was then filtered, washed with MQ-water, dried at 70°C under vacuum condition, and the dry powder is then sieved with a 200 mesh sieve. The end product is named as silica-based chitosan biocomposite material. The biocomposite are labelled based on the composition of the chitosan, successively the composites material prepared from 20 mLof silica with 2% chitosan is labeled as biocomposite SChC1, with 3% chitosan is biocomposite SChC2, with 4% chitosan is biocomposite SChC3, and with 5% chitosan is biocomposite SChC4.

Characterization of the Silica-based ChitosanBiocomposite Material

The characterization of the biocomposite material was carried out by using of XRD, FTIR and SEM. The X-ray diffractometer was used to collect the XRD data at room temperature by using copper K α radiation, and the experimental condition was kept at 40 kV and 30 MA, the scanning rate was maintained of 2° min⁼¹ in range 2 θ from 7° to 70°. The FTIR spectra for the sample was run by using of KBr pellet, scanned in transmission mode with 2 and 4 cm⁼¹ resolution at the range of 4000 to 500 cm⁼¹. Characterization for the surface morphology of the biocomposite material was obtained by using a SEM. Specific surface area, pore radius and pore volume was observed by using GSA, and data analysis was collected by using BET method. Silica-based chitosan biocomposite (0.05-0.2 g) was

Commented [NN9]: It would be better if SChC2 for 2% etc.

degassed (100 <u>e</u>C, 3 hs) in a GSA, followed by weighing and collecting the BET data. The adsorption-desorption isotherm of the biocomposite was measured at 77 K, that was a plot from volume (cc/g) *vs* relative pressure (P/P0).

Adsorption Capacity of Silica-based Chitosan Biocomposite to Cd(II)Ion

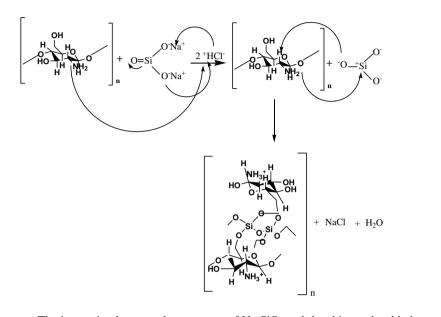
Adsorption capacity of the silica-based chitosanbiocomposites(SChC1, SChC2, SChC3 and SChC4) toward heavy metal ion was studied in solid-phase extraction method. The biocomposite (0.5 g) at different compositions were put into dry columns, followed by addition of 10 mL MQ-water and homogenized at neutral condition (pH 6.0). A solution of 24 ppm Cd²⁺ was loaded into the columnat flow rate 4 ml/min, and the effluent (60 mL) is collected every 15 minutes, and the remaining Cd(II) ion in the effluentwas determined by AAS at λ 228.8 nm.

Results and Discussions

Formation of Silica-based Chitosan BiocompositeMaterial

The silica-based chitosan biocomposite material has successfully been preparedfrom theimmobilization reaction of $Na_2SiO_3(20 \text{ mL})$ with (2-5 %) chitosan by sol-gel method. The chitosan has covalently bounded to the silica surface that was facilitated by the interaction of the protonated amino groups in the organic polymer withthe dissociated of the hydroxyl groups available in the silicabackbone⁴⁰. The reaction mechanism of the formation of silica-chitosanbiocomposite ispresented in equation (1).

Commented [NN10]: Where is gluaradehyde in the reaction scheme?



The interaction between the precursor of Na₂SiO₃ and the chitosan has bind together to form a silica-chitosan composite. Immobilisation reaction has successfully bound the silica and the chitosan to form silica-chitosan biocomposite material. The end product of the silica-chitosan biocomposite material is white crystals, and the weight of the crystal varies depending on the variation amount of the added chitosan in the batch reaction. The weight of the resulted biocomposites materials at different treatment conditions is summarised in Table 1. The results revealed that the weight of the biocomposite material increased significantly as increasing the amount of the chitosan immobilised onto the silica based material. The silica-based chitosan biocomposites are labelled successively as SChC1, SChC2, SChC3 and SChC4 depend on the amount of bounded chitosan in the composite materials.

(1)

Characterization of Silica-based Chitosan Biocomposite Material

The covalent binding between the chitosan and the silica in the immobilization process has been evaluated from the FTIR spectra in the range of 400-4000 cm⁼¹ as presented in Figure 1. It is accertained from the spectra that the mesoporous biocomposite material was containing of amine functional group resulted from the immobilisation of the organic material onto the silica backbone (Figure 1c-f) which can be compared to the spectra of pure silica gel (Figure 1a) and pure chitosan (Figure 1b). The broad bands spectra observed at 790 and 1070 cm⁼¹ correspond to the symmetric Si_O bonds and asymmetric Si_O_Si bonds, and the bands at 1700 and 1550 cm⁼¹ in the FTIRspectra correspond to the amino groups in the

polymer^{38,41}. Furthermore, the bands observed at 1380- cm^{-4} , 1, 310- cm^{-4} , and 1,080 cm⁻¹ are successively corresponding to the presence of C_O_C, C_O, and CH_OH bonds in the biocomposites²⁹.

Characterization of the biocomposite material has been carried out and the XRD patterns are presented in Figure 2. The XRD spectra results showed that the biocomposites (SChC1, SChC2, SChC3 and SChC4) are tend to be amorphous with small crystalline, the crystal area are obtained between $2\theta = 20-24^{\circ}_{-}$ depends on the the amount of immobilised chitosan (Figure 2c-f), which are different to a crystalline structure of the silica at $2\theta = 10.1^{\circ}_{-}$ (Figure 2a) and the the chitosan at $2\theta = 20.3^{\circ}_{-}$ (Figure 2b). The XRD spectra revealed that the amount of the chitosan cointained in the biocomposite suppressed the crystallinity pattern. The polymeric chain in the biocomposite material has proven that the silica and the chitosan were bounded well via covalent bonding^{42,43}.

The SEM imagesforbiocomposites materials and the starting material of silica and chitosan have been comparedas presented in Figure3.It is revealed that the morphologies of the biocomposite materials (SChC1, SChC2, SChC3 and SChC4)(Figure 3c-f) are different to those of starting material of silica gel(Figure 3a) and the chitosan (Figure 3b).The biocomposites(SChC1) possessed as a non-uniform particle, rough surface with with having large pore sice (Figure 3c), while the biocomposite with very high concentration of chitosan(Figure 3f) was tend to have uniform particles^{44,45}.Addition of chitosan to the silica surface resulted to decrease of the total pore volume of the adsorbent because the chitosan occupies the pores in the silica²⁹. Rough surfaces in the biocomposites SChC1 provided more pores to facilitate the absorbadsorbent agent that make it become the best condition to remove heavy metals.

The adsorption/and desorption isotherm for the silica-based chitosan biocompositehas also been evaluated. The adsorption/and desorption isotherm profile for silica-based chitosan biocomposites at various concentration of chitosan in the composite material (SChC1, SChC2, SChC3 and SChC4) have been obtained as presented in Figure 4. The highest adsorption/and desorption profile have been obtained at biocomposites SChC1 and the lowest is found at biocomposite SChC4. It is seen from the results that the adsorption/and desorption of the biocomposites are significantly influenced by the amount of immobilised chitosanthat are bounded in the composite materials^{46,47}. The adsorption/and desorption profile has confirmed that the composite material is assigned to be a mesoporous particle⁴⁸.

Properties of Silica-Based ChitosanBiocomposite Material

The biocomposite with variation concentration of immobilised chitosan onto silica has been evaluated by SAS method followed by BET analysis toward heavy metal. The properties of the developed biocomposites based on the mass, BET measurements, and the absorption adsorption capacity is summarised in Table 1. The mass of biocomposite increased significantly with addition of higher concentration of the chitosan. This reveals that immobilisation the chitosan has completely immobilised to the silica main body. It is observed that more chitosan bounded onto the silica surface resulted to decrease of total pore volume of adsorbent, this might bedue to the occupation of the pores of silica by the chitosan.The BET analysis has showed that the surface area of the biocomposite varies dependingon the amount of chitosan added in the biocomposite material. The surface area for the biocomposites are smaller compare to the surface area of the chitosan raw material (see data in Table 1). The observations showed that the more the amount of chitosan is added, the smaller the specific surface area, but this pattern does not occur with the addition of 5% chitosan (SChC4). This may be in line with the morphological observations in SChC3 when mixingsilica and 4% chitosan is not very homogeneous, so the aggregation occurs as chitosan buildup occurs on a part of the silica surface so that chitosan is bound unevenly to the silica surface and causes the specific surface area is very significantly different from SChC4.

Capacity Abd sorption Capacity of Silica-Based Chitosan Biocomposite

The adsorption characteristic of the biocomposite with variation of the chitosan in the material to remove Cd(II) ion is examined. The adsorption profile of the chitosan-based silica biocomposite was recorded from the concentration of Cd²⁺ after loading (Ct) compare to the original 24 ppm Cd^{2+} (C₀) as presented in Figure 5, and the <u>adsorption</u> capacity of the biocomposite materials are listed in Table 1. The variation in the immobilised chitosan resulted to the variation of the BET values, influenced the active sites of the material, and promote adsorbtion capacity of biocomposite material to a target ion43,48,49. The biocomposites are high in the adsorption capacity with time. The adsorption profiles of the biocomposite are almost similar from one to another but different in the adsorption capacity in related to the variation of the immobilised chitosan in the biocomposite. The highest adsorption was obtained in SChC1 with adsorption capacity of 16.16 mg/g Cd^{2+} . High concentration of chitosan in the biocomposite did not improve adsorption ability toward metal ion. The capacity adsorption decrease with time which is able to adsorb the ion up to 75 minutes, followed by

releasing the ion until obtaining an equilibrium after 120 minutes. The <u>absorptionadsorption</u> capacity of the chitosan-based silica biocomposites varies with different concentrations of immobilised chitosan (see results in Table 1). The <u>absorptionadsorption</u> capacity is confirmed to be related to the pore size and area where the biggest <u>absorptionadsorption</u> capacity was observed in a very small size of the composite particle. The chitosan-based silica biocomposite of SChC1 was the best <u>absorbadsorb</u>ent material with having high ability to load metal ion in batch condition.

Conclusion

The silica-based chitosan biocomposite has successfully fabricated through the mixture of precursorNa₂SiO₃ and chitosan solutionby using sol-gel method. The interaction between the silica and the chitosan has bind together by the crosslinking agent of glutaral dehyde to form a silica-based chitosan biocomposite. The variation in the concentration of the chitosan that are immobilised into the silica backbone resulted in different properties of the biocomposite material. The biocomposite material prepared from 20 mL silica and 2% chitosan (SChC1) is assigned to be the best silica-based chitosan biocomposite. The biocomposite material SChC1 is amorphous with small crystalline, non-uniform particle, rough surface with with having large pore sice area between $2\theta = 20-24^{\circ}$. The biocomposite has high absorptionadsorption capacity towards metals ion which is potential to be applied in the production of low cost adsorbent material.

Acknowledgment

The authors acknowledged the financial support from The Ministry of Research, Technology and Higher Education of the Republic of Indonesia, Directorate Research and Community Service, Directorate General Strengthen Research and Development, Financial Year 2018, under Penelitian Disertasi Doktor, Contract No. 027/UN33.8/LL/2018. The authors also acknowledge Dr. Isli Iriani Pane from the Language Centre of Universitas Negeri Medan who have helped in the English proofreading process.

References

- 1 Hu Z, Zhang D Y, Lu S T, Li P W & Li S D, Mar. Drugs. 16(2018) 273.
- 2 Marpu S B & Benton E N, Int. J. Mol. Sci.19(2018) 1795.
- 3 Rodríguez-Vázquez M, Vega-Ruiz B, Ramos-Zúñiga R, Saldaña-Koppel D A & Quiñones-Olvera L F, *BioMed Res. Int*. (2015) Article ID 821279.
- 4 Szymańska E & Winnicka K, Mar. Drugs13(2015) 1819.

- 5 Elgadir M A, Uddin M S, Ferdosh S, Adam A, Chowdhury A J K & Sarker M J I, *J. Food Drug Anal*.23 (2015) 619.
- 6 Kyzas G Z & Bikiaris D N, Mar. Drugs13(2015) 312.
- 7 Nurwahyuni I & Sinaga R, Pak. J. Bot. 50(2) (2018) 667.
- 8 Nurwahyuni I, Proceeding of 6thAnnual International Conference on Advances in Biotechnology (BIOTECH 2016) April 2016, Singapore, (2016) pp. 35-39.
- 9 Nurwahyuni I & Sinaga R, Int J Pharm Bio Sci 5(4)(B) (2014) 863.
- 10 Wang J, Yang S, Li C, Miao Y, Zhu L, Mao C & Yang M, ACS Appl Mater Interfaces 9(27) (2017) 22259.
- 11 Kumaravel V, Damodaran K, Gandhi S, & Krishnan U M, Bull. Mater. Sci. 41(2018) 133.
- 12 Simatupang L, SiburianR, SitanggangP, DoloksaribuM, SitumorangM, &Marpaung H, *Rasayan J. Chem.* 11(2) (2018) 819.
- 13 Yazdani M, Bahrami H & Arami M, Sci. World J(2014) Article ID 370260.
- 14 Prabhu S M, & Meenakshi S, DesalinWater Treat.53 (13) (2015) 3592.
- 15 Tamburaci S & Tihminlioglu F, J Mater Sci: Mater Med 29 (2018) 1.
- 16 Maleki A, Haji R F, Ghassemi M & Ghafuri H, J. Chem. Sci. 129(4) (2017) 457.
- 17 Venkatesan J, Bhatnagar I & Kim S K, Mar. Drugs12(2014) 300.
- 18 Hu J, Liu W, Liu H, Wu L & Zhang H, *Materials*11(2018) 1881.
- 19 Badwan A A, Rashid I, Al Omari M M H & Darras F H, Mar. Drugs13(2015) 1519.
- 20 Saravanakumar A & Dharmendirakumar M, Indian J Chem Technol 25(6) (2018) 546.
- 21 Sihombing K, Tamba M C, Marbun W S & Situmorang M, Indian J Chem.57A(2) (2018) 175.
- 22 Situmorang M & Nurwahyuni I, Indones. J. Chem. 17(3)(2017) 461.
- 23 Situmorang M, Gooding J J, Hibbert D B & Barnett D, *Electroanalysis* 13(18) (2001) 1469.
- 24 Situmorang M, Gooding J J, Hibbert D B & Barnett D, Biosens. Bioelectron. 13(9) (1998) 953.
- 25 Teepoo S, Dawan P & Barnthip N, Biosensors7(2017) 47.
- 26 Rao D, Sheng Q & Zheng J, J. Chem. Sci. 128(9) (2016) 1367.
- 27 Ang L F, Por L Y & Yam M F, PLoS ONE 8(8) (2013) e70597.
- 28 Lee E J, Jun S H, Kim H E, Kim H W, Koh Y H & Jang J H, *J Mater Sci: Mater Med* 21 (2010) 207.
- 29 Djelad A, Morsli A & Robi M, Molecules 21(2016) 109.
- 30 Zhang J, Zhang Y, Li R & Pan Q, Indian J Chem Technol 19(1) (2012) 161.
- 31 Gonçalves M C, Molecules 23(2018) 2021.
- 32 Baino F, Fiorilli S and Vitale-Brovarone C, *Bioengineering*4(2017) 15.
- 33 Bhullar N, Kumari K, &Sud D, Indian J Chem Technol, 24(5)(2017)560.

- 34 Budnyak T M, Pylypchuk I V, Tertykh V A, Yanovska E S & Kolodynska D, Nanoscale Res. Lett. 10 (2015) 87.
- Li J, Cai C, Li J, Li J, Li J, Sun T, Wang L, Wu H & Yu G, Molecules 23(2018) 2661.
- 36 Kong J L, Chang J S & Chang K L B, J Nanopart Res 15 (2013) 1945.
- 37 Budnyak T M, Yanovska E S, Kichkiruk O Y, Sternik D & Tertykh V A, *Nanoscale Res. Lett.* 11 (2016) 492.
- 38 GandhiMR& Meenakshi S, Int. J. Biol. Macromol. 50(3)(2012) 650.
- 39 Shchipunov A Y, Karpenko Y T, Bakunina Y I, Burtseva V Y & Zvyagintseva N T,J. Biochem. Biophys Methods 58 (2004) 25.
- 40 da Silva S R, de Albuquerque N J A, de Almeida R M &de Abreu F C, *Materials*10 (2017) 1132.
- 41 Wei P R, Kuthati Y, Kankala R K & Lee C H, Int. J. Mol. Sci 16(2015) 20943.
- 42 Zhang B, Hu R, Sun D, Wu T & Yujiang, Sci. Rep.8 (2018) 15397.
- 43 Chernev G, Todorova E, Djambazov S, Salvado I M M & Ivanova J, *J. Chem. Technol. Metall*.49(2)(2013) 128.
- 44 Zhou D, Qi C, Chen Y X, Zhu Y J, Sun T W, Chen F & Zhang C Q,*Int. J. Nanomed* 12 (2017) 2673.
- 45 Nithyaa R, Gomathi T, Sudha PN, Venkatesan J, Anil S, and Kim SK, *Int. J. Biol. Macromol.* 87 (2016) 545.
- 46 Dalagan J Q & Enriquez E P, Bull. Mater. Sci. 37(3) (2014) 589.
- 47 Budnyak T, Tertykh V & Yanovska E, Mater Sci-Medzg. 20(2) (2014) 177.
- 48 Viswanatha N, Pandi K & Meenakshi S, Int J Biol Macromol. 70 (2014) 347-353.
- 49 Prabhu S M & Meenakshi S, Desalin Water Treat. 53(13) (2015) 3592.

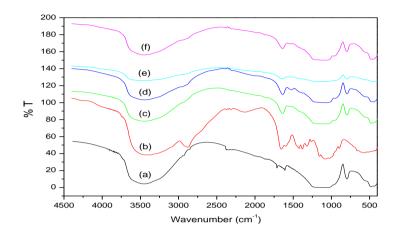


Fig. 1. The FTIR spectra of: (a) pure silica gel (Sg), (b) pure chitosan (Ch); and for silica-based chitosan biocomposite of: (c) SChC1 (20 mL : 2%), (d) SChC2 (20 mL : 3%), (e) SChC3 (20 mL : 4%), and (f) SChC4 (20 mL : 5%).

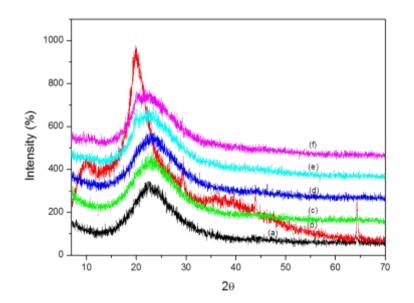


Fig. 2. The XRD spectra of: (a) pure silica gel (Sg), (b) pure chitosan (Ch); and for silica-based chitosan biocomposite of: (c) SChC1 (20 mL : 2%), (d) SChC2 (20 mL : 3%), (e) SChC3 (20 mL : 4%), and (f) SChC4 (20 mL : 5%).

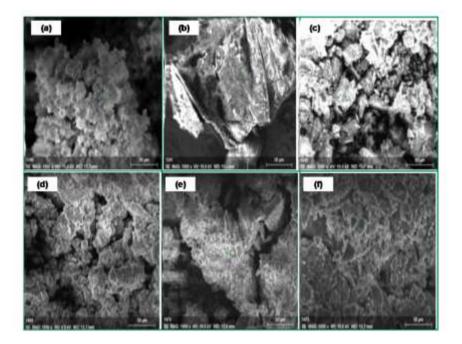


Fig. 3. The SEM images of: (a) pure silica gel (Sg), (b) pure chitosan (Ch); and for silica-based chitosan biocomposites of: (c) SChC1 (20 mL : 2%), (d) SChC2 (20 mL : 3%), (e) SChC3 (20 mL : 4%), and (f) SChC4 (20 mL : 5%).

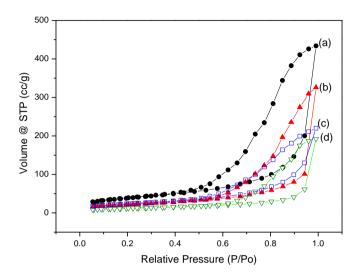


Fig. 4. The adsorption-desorption isotherm for silica-based chitosan biocomposites of: (a) SChC1 (20 mL : 2%), (b) SChC2 (20 mL : 3%), (c) SChC3 (20 mL : 4%), and (d) SChC4 (20 mL : 5%). The measurements are made at 77 K.

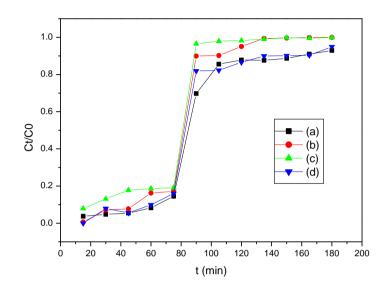


Fig. 5. The adsorption capacity toward Cd(II) ion by solid-phase extraction method for biocomposite materials of: (a) SChC1 (20 mL : 2%), (b) SChC2 (20 mL : 3%), (c) SChC3 (20 mL : 4%), and (d) SChC4 (20 mL : 5%). The data are recorded from the proportion of the amount of Cd^{2+} after loading (C_t) compared to the original Cd^{2+} (C₀) for 18 minutes.

Starting material	Mass	The BET measurement			Absorption
and the label of biocomposite materials ⁺	(g)	Surface area (m²/g)*	Average pore size (A)*	Total pore volume (cc/g)*	Adsorption capacity (mg/g) [#]
Silica gel	na	374.994	1.5469 x 10 ¹	2.900 x 10 ⁻¹	na
SChC1 (20 mL:2%)	0.784	138.509	9.69225 x 10 ¹	6.712 x 10 ⁻¹	16.16
SChC2 (20 mL:3%)	0.971	76.644	1.31547 x 10 ²	5.041 x 10 ⁻¹	13.76
SChC3 (20 mL:4%)	1.066	42.079	1.41445 x 10 ²	2.976 x 10 ⁻¹	12.48
SChC4 (20 mL:5%)	1.176	85.021	8.00543 x 10 ¹	3.403 x 10 ⁻¹	14.93

Table 1. The properties of the silica-based chitosan biocomposites materials based on the mass, BET data, and the absorptionadsorption capacity toward Cd(II) ion.

* Biocomposite material is prepared from 20 mL silica with (2% - 5%) chitosan.
* Calculated by using Brauner, Emmet and Teller (BET) method.
Calculated from the concentration of Cd(II) ion after loading (Ct) compare to the original Cd(II) ion (C0)